A Theoretical Framework for Performance Characterization of Elastography: The Strain Filter

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Abstract—This paper presents a theoretical framework for performance characterization in strain estimation, which includes the effect of signal decorrelation, quantization errors due to the finite temporal sampling rate, and electronic noise. An upper bound on the performance of the strain estimator in elastography is obtained from a strain filter constructed using these limits. The strain filter is a term used to describe the nonlinear filtering process in the strain domain (due to the ultrasound system and signal processing parameters) that allows the elastographic depiction of a limited range of strains from the compressed tissue. The strain filter predicts the elastogram quality by specifying the elastographic signal-to-noise ratio \( SNR_e \), sensitivity, and the strain dynamic range at a given resolution. The dynamic range is limited by decorrelation errors for large tissue strain values, and electronic noise for low strain values. Tradeoffs between different techniques used to enhance elastogram image quality may also be analyzed using the strain filter.

I. INTRODUCTION

ULTRASONIC TECHNIQUES for measuring the elasticity of compliant tissue generally rely on the estimation of the strain \([1]–[5]\). Elastography, a technique of estimating the axial strain using differential displacements of the tissue elements due to tissue compression, was proposed by Ophir et al. \([1]\).

This paper introduces the strain filter concept for the performance characterization of the strain estimator in elastography. The behavior of the upper bound of the \( SNR_e \) as a function of axial tissue strain forms a bandpass filter in the strain domain. The strain domain refers to the entire range of strains present in the compressed tissue. The strain filter consists of a graphical and analytical representation of the allowable range of strain values and their resulting \( SNR_e \) for a given elastographic resolution. The width of the strain filter specifies the dynamic range, and its height the respective \( SNR_e \) value of the estimated strain.

A block diagram elucidating the strain filter concept is presented in Fig. 1. Elastography uses small mechanical compressions on soft biological tissue that have a wide range of elastic moduli. A distribution of strains in the medium is caused by the quasi-static compression under certain mechanical boundary conditions. The input to the strain filter is the actual tissue strain characterized by an infinite dynamic range and \( SNR_e \), fine (denoted by \( \epsilon \) where epsilon is a very small number) resolution and sensitivity. However, the interaction of the actual tissue strain with the ultrasound system and signal processing parameters corrupts the elastogram obtained, in the sense that it now has a finite dynamic range, \( SNR_e \), sensitivity, and resolution. The strain filter predicts the elastographic image quality (\( SNR_e \), sensitivity, and dynamic range at a given resolution) in terms of the signal processing and system parameters used to obtain the elastogram. The strain filter therefore provides a quantitative assessment of the elastogram quality, in terms of the four parameters described above.

In addition, the strain filter makes it possible to select the appropriate ultrasound system and signal processing parameters to obtain the best possible elastogram under given tissue conditions. In subsequent sections of this paper we discuss the effects of the various ultrasound system parameters such as pulse center frequency and band-
width, and signal processing parameters (window size and overlap factor). Tradeoffs among different parameters and techniques that enhance elastogram quality may be evaluated and predicted using the strain filter. As illustrated in Fig. 1, techniques such as temporal stretching (TS) [6]–[8], multicompression averaging (MA) [5],[9],[10], and stretching multicompression averaging (SMA) [9],[10], reduce signal decorrelation, i.e. improve the correlation coefficient, thereby enhancing the performance of the strain filter. The improvements in the elastogram obtained using these techniques may be quantitatively predicted using the strain filter.

The ideal strain filter therefore has an infinitely high, flat all-pass characteristic shape in the strain domain, which means that all local tissue strains are displayed in the elastogram with infinite $SNR_e$; it also means that the strain dynamic range in the elastogram is infinite as well. Practical strain filters, however, have a bandpass characteristic shape in the strain domain, where the $-3$ dB width of this bandpass characteristic may be defined as the elastographic dynamic range. Under typical conditions, a $-3$ dB dynamic range of $\approx 30$ dB is predicted by the strain filter; this is consistent with the range of strains measured experimentally using a single compression.

A description of the literature used to develop the strain filter is presented in Section II. The theoretical model for the strain filter is developed in Section III. Simulations to validate the theoretical model are presented in Section IV. The contributions of this paper are summarized in Section V.

II. BACKGROUND

Development of the strain filter concept is based on obtaining the tightest bound on the variance of the strain estimator that includes all noise sources. Since the strain estimator uses time-delay (displacement) estimation to compute strain, the vast literature on time-delay estimation can be adapted for the strain estimation problem.

A. Noise Sources

The Cramér-Rao lower bound (CRLB) is the most commonly used lower bound on time-delay variance [13]–[16]. The CRLB, however, can be achieved only for large post integration SNR [13],[14] and zero strain, conditions that are unrealistic for strain estimation. The expression of the CRLB derived by Walker and Trahey [17] (for partially correlated signals) increases the bound on the variance to a more achievable level. While electronic and quantization noise contributions (sonographic SNR, denoted by $SNR_S$) are accounted for in the expression for the CRLB through the SNR term [13]–[16], the added effect of decorrelation on the variance of the time delay estimate has been modeled recently by Walker and Trahey [17]. The authors also indicate that TDE performance can be worse than the CRLB at poor SNRs ($<15$ dB) and low correlation coefficient values ($<0.5$).

Signal decorrelation, a significant source of error in the displacement estimate, increases rapidly with tissue compression. Decorrelation errors are caused by the relative displacement of the scatterers in all three dimensions due to tissue compression. In this paper, signal decorrelation is modeled as a noise process that reduces the composite signal-to-noise ratio ($SNR_C$) in the echo-signal. $SNR_C$ is a combination of the constant electronic noise level and a varying component due to signal decorrelation. To incorporate the contributions due to signal decorrelation it is necessary to convert the correlation coefficient to an SNR measure ($SNR_p$). A relationship between $SNR_p$ and the correlation coefficient has been independently derived by Friemel [18], and by Céspedes et al. [12]. The expression used to obtain $SNR_C$ is presented in Appendix A.

Increasing signal decorrelation errors with tissue strain reduce $SNR_C$, causing the strain estimation variance to exceed the CRLB (as also noted by Walker and Trahey [17]), thereby necessitating the need for a more advanced lower bound to predict the variance of the strain estimator. The modified Ziv-Zakai lower bound proposed by Weinstein and Weiss [13],[14] is used in this paper to obtain an accurate lower bound.

B. The Ziv-Zakai Lower Bound (ZZLB) on Time-Delay Estimation

Weinstein and Weiss present plots of the lower bound of the time delay variance versus the post integration SNR [13],[14] (defined as a product of the bandwidth, data window length, and the $SNR_C$), that divides the post integration SNR domain into three distinct regions (low, moderate, and high). Since $SNR_C$ is the only parameter in the expression for the post integration SNR that varies with tissue strain, its value determines the appropriate lower bound.

The Cramér-Rao lower bound (CRLB) is applicable only to high post integration SNR situations (at low strains). In this region, time-delay estimation is subject only to local errors (ambiguity-free mode of operation). At moderate SNR values, the lower bound exceeds the CRLB and obeys the Barankin bound. In this region, ambiguities in the signal phase cannot be resolved; however, an estimate of the time-delay estimate may still be obtained using the correlation between signal envelopes. At low post integration SNR values, the lower bound approaches a constant level. In this region, both envelope and phase ambiguities exist, and the time delay cannot be estimated correctly. The thresholds separating these three regions are determined by the value of the post integration SNR [13],[14]. The above combination of lower bounds on the variance of the time-delay estimator is referred to as the Ziv-Zakai lower bound [13],[14]. The ZZLB provides the tightest bound on the variance of the time-delay estimator. The modified ZZLB and threshold values at the transition points are discussed in Appendix B.
In this paper, the strain variance is computed from the TDE variance using the expression derived by Walker and Trahey [17], as long as the ZZLB coincides with the CRLB. The decay in the value of the correlation coefficient with tissue compression has been modeled using an analytic expression derived by Meunier and Bertrand in [19] to estimate decorrelation effects due to rotation, translation, and biaxial deformation of the elastic tissue elements. The theoretical development of the strain filter is presented in the next section.

III. DEVELOPMENT OF THE STRAIN FILTER

Axial strain $s$ is the displacement gradient, which may be estimated from two adjacent time delay estimates separated by a time interval $\Delta t$ [1], assuming a constant speed of sound in the tissue, viz:

$$s = \frac{\tau_2 - \tau_1}{\Delta t},$$

(1)

where $\tau_1$ is the time delay estimate at time $t$, and $\tau_2$ is the time delay estimate at time $t + \Delta t$ for a data window with duration $T$.

Since the strain estimate is obtained from a linear combination of two random variables (time delay estimates separated by $\Delta t$), the variance of the strain estimator depends on the variance of the time delay estimator. Assuming stationarity, the variance of the strain estimate ($\sigma^2_s$) is expressed in terms of the variance of the time delay estimates ($\sigma^2_T$) in [20], and is given by:

$$\sigma^2_s \geq \frac{2\sigma^2_T}{T\Delta t}.$$  

(2)

(2) illustrates that for a given window size and overlap, strain variance is reduced when the variance in the time delay estimate is minimized. The resolution in the elastogram is reduced with an increase in $T$. Large overlapping windows also generate correlated errors which bias the strain estimate. In addition, an optimal window size exists, where the strain estimation variance is minimum, with an increase in the variance observed as the window size is increased or decreased [6, pp. 122, 21]. Therefore the strain estimation variance cannot simply be reduced in the limit as $T \rightarrow \infty$, since $\sigma^2_T$ also increases.

A. The Strain Filter in Elastography

A measure of elastographic image quality was described [6],[7] in terms of the mean to standard deviation ratio ($SNR_e$) of the elastogram:

$$SNR_e = \frac{\mu_s}{\sigma_s},$$

(3)

where $\mu_s$ and $\sigma_s$ are, respectively, the mean and standard deviation of the strain estimates in a region of uniform elasticity. The upper bound of the $SNR_e$ is obtained when the total tissue strain ($s_t$) and the lower bound on the strain estimation standard deviation ($\sigma_{ZZLB}$) are substituted in (3):

$$SNR_e^{UB} = \frac{s_t}{\sigma_{ZZLB}}.$$  

(4)

Incorporating the modified ZZLB expression for the TDE variance (see Appendix B) into $\sigma_{ZZLB}$ using (2), we obtain:

$$\sigma^2_{ZZLB} \geq \begin{cases} 
\frac{\mu^2}{\delta^2 T\Delta t}, & BSNR_C < \gamma' \\
\frac{2\mu^2}{T\Delta t}, & \delta' < BSNR_C < \mu' \\
\frac{2\sigma^2_{BB}}{T\Delta t}, & \eta' < BSNR_C 
\end{cases}$$

(5)

where $\gamma'$, $\mu'$, $\delta'$, and $\gamma'$ are the modified thresholds (B-2) scaled by the factor $\frac{2}{T\Delta t}$. (5) shows the three distinct operating regions for $\sigma^2_{ZZLB}$, depending on the value of $BSNR_C$. A distinct threshold region is observed between the CRLB and the Barankin bound; however, the variance increases exponentially in this threshold region (see Appendix B). Accurate estimation of the strain is possible only within the CRLB.

The minimum variance of the time delay estimator is given by the CRLB [13]-[17]. The CRLB for time delay estimation has been adapted for partially correlated signals by Walker and Trahey in [17], and is given by:

$$\sigma^2_{CRLB} \cong \frac{3}{2\pi^2T (B^3 + 12Bf_o^3)} \left[ \frac{1}{\rho^2 (1 + \frac{1}{SNR^2})^2 - 1} \right]$$

(6)

where $f_o$ is the center frequency, $B$ is the bandwidth, $\rho$ is the correlation coefficient, and the SNR term represents only the contribution due to electronic noise ($SNR_s$). This closed-form expression obtained for signals with a rectangular spectrum assumes that the variance of the time delay estimate is bounded by the CRLB. The Barankin bound exceeds the CRLB by a factor of $12 \left( \frac{f_o}{B} \right)^2$ [13],[14], and is given by:

$$\sigma^2_{BB} = 12 \left( \frac{f_o}{B} \right)^2 \sigma^2_{CRLB}$$

(7)

The lower bound on the variance of the strain estimate ($\sigma^2_{ZZLB}$) is obtained by substituting the bound on the variance of the TDE obtained using (6) and (7) into (5).

The variation of $SNR_e^{UB}$ (4) with tissue strain is defined as the strain filter. Three distinct regions constitute the strain filter, which depends on the appropriate lower bound that contributes to $\sigma^2_{ZZLB}$ (5).

B. Effect of Decorrelation Due to Strain on the Correlation Coefficient

Decorrelation errors increase with tissue strain, causing a decay in the values of the correlation coefficient. The correlation coefficient with motion compensation due to axial deformation of elastic tissue for a 2-D Gaussian model
has been derived by Meunier and Bertrand in [19], and is given by:

$$\rho = \frac{2\sqrt{\alpha\beta}}{\sqrt{2(\alpha^2 + 1)(\beta^2 + 1)}} e^{-\frac{1}{2}\left(\frac{f}{\alpha c}\right)^2\left(\frac{1}{\beta} - \frac{1}{2}\right)^2}$$

(8)

where $f$ is the spatial frequency in cycles/mm ($f = \frac{2f_c}{c}$ where $c$ is the speed of sound in tissue = 1.54 mm/s), and $\sigma_f$ is the standard deviation of the Gaussian envelope in cycles/mm ($\sigma_f = \frac{2\pi}{2\pi \sigma_f}$, and $\sigma_f = \frac{1}{2\pi \sigma_f}$, where $\sigma_f$ is the spatial standard deviation and $\sigma_f$ is the standard deviation of the Gaussian envelope in the frequency domain), $\alpha$ represents the axial compression where $\alpha = 1 - s$, and $s$ is the tissue strain. The corresponding lateral expansion is denoted by $\beta$ (with the incompressibility constraint $\alpha\beta = 1$). The value of the correlation coefficient is substituted in the expression for the CRLB (6) to model decorrelation effects.

Expressions for the CRLB [13]–[17] have been derived for flat bandlimited signal and noise spectra. However, the correlation coefficient in [19] is derived for a Gaussian-shaped spectrum rather than a rectangular spectrum. A reasonable approximation was obtained by Céspedes et al. [11] using a rectangular spectrum centered at the Gaussian center frequency with the same mean square amplitude value as the Gaussian spectrum. The equivalent noise spectral bandwidth [22, pp. 141] is defined by:

$$B = \frac{\int_0^{\infty} P(f) df}{P(f)_{\text{max}}} = \sqrt{2\pi \sigma_f}$$

(9)

where $B$ is the bandwidth of a rectangular spectrum with the same total power and peak amplitude as the Gaussian pulse spectrum $P(f)$.

Using the following typical signal parameters, $T = 1$ mm (1.3 $\mu$s), $f_o = 5$ MHz, $B = 3$ MHz (60% bandwidth), $SNR_b = 100$ (40 dB), and interval between strain estimates $\Delta z = 0.5$ mm ($\Delta t = 0.66$ $\mu$s). The lower bound on the standard deviation of the strain estimator (normalized by its value at 8% strain) is plotted along with the corresponding correlation coefficient in Fig. 2, for increasing strain values. Note from Fig. 2 that the lower bound on the standard deviation ($\sigma_{\text{ZZLB}}$) increases dramatically for strain values $>0.5%$. This has been previously noted by O’Donnell et al. [5] and Céspedes [6, pp. 115]. They show that the standard deviation of the strain for a 1-D simulation increases dramatically due to decorrelation associated with large strain values ($>3\%$) for a fixed correlation integration time. For the same reason, past work in elastography has used small ($\leq 2\%$) applied strains [1],[2].

Decorrelation errors increase with tissue strain, reducing the value of the correlation coefficient, and causing $\sigma_{\text{ZZLB}}$ to move from the CRLB to the Barankin bound or the constant variance level as shown in (5). The three distinct regions in (5) are observed in plots of the strain filter as shown by the curves in Fig. 3. Figure 3 shows the strain filter obtained for a 3 MHz rectangular bandwidth using the ZZLB (CRLB for strains $\leq 10\%$, Barankin bound for strains $>10\%$ and $<30\%$, and the constant variance level for strains $>30\%$). As the lower bound coincides with the Barankin bound, the performance of the strain filter drops sharply, with a further drop in performance observed as the variance coincides with the constant variance level. Note that the strain filter obtained using the ZZLB has a smaller dynamic range than the strain filter obtained under the optimistic assumption that the strain estimation variance is always bounded by the CRLB.

The range of strains that can be reliably estimated using the elastogram determines the dynamic range of the strain filter. The dynamic range of the strain estimator in decibels is defined as follows:

$$DR = 20 \log \left[ \frac{s_{\text{max}}}{s_{\text{min}}} \right],$$

(10)

where $s_{\text{max}}$ is the maximum strain and $s_{\text{min}}$ is the minimum strain at a specified $SNR_c$ level in the strain filter. The quantity $s_{\text{min}}$ also defines the sensitivity of the strain filter. A 1% tissue strain corresponds to a decibel value of $-40$ dB. The dynamic range estimated for a $-3$ dB cutoff level of the strain filter ($SNR_c$ level of 10) predicts a 30 dB dynamic range (observed from Fig. 3) for the strain filter obtained using the ZZLB bound, compared to the 40 dB dynamic range predicted using the CRLB. However, the dynamic range obtained experimentally for a single compression agrees closely with the dynamic range predicted using the ZZLB.

The slope of the strain filter for low strain values is determined primarily by electronic noise contributions, with decorrelation noise contributing to the progressive flattening of the curve. The plateau in the strain filter is caused by increasing decorrelation errors; however, the variance of the strain estimate is still bounded by the CRLB in this region. At high strain values, the precipitous drop in
Fig. 3. The strain filter illustrating the distinct regions of strain estimation obtained using (5), along with the strain filter obtained under the optimistic assumption that the strain estimation variance is always bounded by the CRLB adapted for partially correlated signals. Note that the dynamic range of elastography may be determined from the width of the strain filter, and the heights give the respective $SNR_e$ at every strain level.

The shape of the strain filter also depends on the variation of $T$, $f_0$, and $B$ terms in the denominator of (6). For example, the performance of the strain filter for different values of the bandwidth is shown in Fig. 4. With an increase in the system bandwidth, there is an improvement in the maximum $SNR_e$ value, sensitivity, and dynamic range of the strain filter. In other words, signal decorrelation reduces with an increase in the bandwidth. The strain filters obtained at different center frequencies are presented in Fig. 5. Note the improvement in the sensitivity, dynamic range, and $SNR_e$ obtained with increasing pulse center frequency. The maximum attainable $SNR_e$ is approximately proportional to the square of the center frequency.

The resolution of elastography depends on $T$ and $\Delta t$, and is limited only by the correlation length of the ultrasound system. The resolution in the elastogram improves with a decrease in the value of $T$ or an increase in system bandwidth. All the performance plots of the strain filter are plotted for a constant value of the resolution. A family of performance curves can be obtained at different resolution levels. In general, as resolution increases, the maximum value of the $SNR_e$ and the dynamic range of the strain filter decrease. The dynamic range, sensitivity, and $SNR_e$, along with the resolution, provide a complete characterization of the noise properties in the performance of the strain estimator in elastography.

Simulation experiments in the next section are used to confirm the validity of the theoretical strain filter model developed in this section.

IV. SIMULATION

A simulation experiment to quantify the performance of the strain estimator for a 1-D tissue mechanical model is presented in this section. The 1-D model used in the simulation accounts only for the decorrelation of the echo-signal due to the axial component of the strain. The theoretical model can be adapted for the 1-D case by setting $\beta = 1$ in (8). In the simulation of the ultrasound system, a 1-D sampled array is used to insonify a 2-D point scatterer medium.

A. Method

The pre- and post-compression echo-signals are generated using a transducer with a center frequency of 5 MHz and $-3$ dB bandwidth of 3 MHz (pulse standard deviation of $0.27 \mu s$). The A-scan was sampled at 50 MHz. Cross-correlation analysis was performed using a 1 mm $(1.3 \mu s)$ overlapping window with 0.5 mm $(0.66 \mu s)$ overlap between consecutive windows.

The transducer is modeled as a 1-D sampled aperture composed of point subtransducer elements equally spaced...
by $\lambda/2$. Each subtransducer element is modeled as a point source or receiver with a two-way Gaussian transfer function. The scattering medium is modeled as a 2-D array of point scatterers. The scatterer density in the media was set to 48 scatterers/pulse width. The elastic target is assumed to have a Poisson’s ratio $\approx 0$. This implies that lateral and elevational decorrelation effects due to scatterer motion are ignored. The applied stress is assumed to propagate uniformly so the localized stress is constant throughout the medium. The displacement of each scatterer is a function of the applied strain, and is modeled by considering an equivalent 1-D spring system described in [6]. The spring constant is a function of the Young’s modulus of the tissue. The applied strain is the same as the tissue strain for a uniformly elastic homogenous medium.

The pre-compression A-line is obtained from the randomly distributed scatterers. Each scatterer location is then changed depending on the compressive force and a post-compression A-line generated. The stretched post-compression A-line is generated by applying a linear stretch factor on the post-compression signal. The process is repeated for 28 different lateral locations in the simulated phantom to obtain independent A-line pairs (lateral step $>\text{beamwidth of the transducer}$). The strain values are computed from the individual A-line pairs. Time-delay estimation is performed using the normalized cross-correlation function, with the strains computed using (1).

B. Results

Fig. 6 shows the mean $SNR_e$ value and its standard deviation (error bars), derived from 28 independent simulations, before and after temporal stretching. The x-axis represents the total applied compressive strain expressed as a percentage; $SNR_e$ is plotted, along the y-axis. The theoretical curve of the strain filter for the 1-D case is also shown in the figure.

Observe from Fig. 6 that both the strain filter curves obtained from the simulation are completely bounded by the theoretical strain filter curve. The theoretical strain filter curve forms the upper bound, which determines the attainable experimental or simulation performance. For strain values $>2\%$, decorrelation errors cause the drop in the value of the estimated strain. The CRLB bound on the variance is no longer applicable for strain values $>2\%$, as shown by the simulation experiment, since signal decorrelation introduced due to tissue compression dramatically increases the variance in the strain estimate.

Inaccurate estimation of the time-delay (increased variance) is primarily due to the detection of false peaks and jitter [17]. The simulation results presented in this section are obtained using the basic normalized correlation coefficient function. Non-linear processing to remove phase ambiguities (contributing to the reduction of false peaks [17]) was not performed. Sophisticated algorithms that incorporate additional processing to remove false peaks can therefore significantly improve the performance of the strain estimator.

V. Conclusion

This paper presents a theoretical framework for quantitative assessment of the quality of elastograms. This framework is described as a strain filter that is typically a bandpass filter in the strain domain. This filter allows only a restricted range of strain values to be included in the elastogram. The deviation of the strain filter from an ideal all-pass characteristic in the strain domain is due to the ultrasound system parameters, the finite value of the sono-
graphic SNR, and the effects of signal decorrelation. Signal decorrelation determines the largest value of strain that is accurately estimated, while $SNR_S$ determines the smallest measurable strain value. The dynamic range of the system is thus limited on the low end by electronic noise effects, and on the high end by signal decorrelation, resulting in a bandpass filter in the strain domain.

The range of strains over which the CRLB specifies the variance of the time delay defines the optimum performance range for the strain estimator. The formulation of the CRLB by Walker and Trahey does not address the implications of decorrelation causing the deviation of the time delay variance from the CRLB for large tissue strains. Application of the $\rho - SNR_\rho$ relationship developed by Friemel [18] and Céspedes et al. [12] clearly demarcates the regions over which the CRLB, the Barankin bound, and the constant variance level are applicable. Signal decorrelation may be reduced using a combination of small compressions that allow successful temporal stretching, (e.g., temporal stretching described by Céspedes and Ophir [6]–[8], and SMA described by Huang et al. [9], [10]). Temporal stretching is currently used to correct for decorrelation only along the axial direction. Lateral and elevational decorrelation errors are more difficult to correct, since the scatterers move out of the beam for large strains. Multicompression averaging in conjunction with temporal stretching of the strain estimates obtained from several small compressions, increases both the $SNR_e$ and the dynamic range of strain estimation without sacrificing resolution. The improvements in the elastogram obtained using these techniques may be quantitatively predicted using the theoretical strain filter model developed in this paper.

The strain filter concept developed in this paper provides a graphical framework for characterizing elastography. The strain filter quantified the elastogram image quality ($-3$ dB dynamic range $30$ dB using the ZZLB and maximum elastogram $SNR_e \approx 23$ dB for a single compression), for the signal parameters specified in Section III. In a similar manner, different elastographic signal processing techniques can now be compared and their performances quantified using the dynamic range, sensitivity, resolution, and maximum $SNR_e$ obtained from the strain filter. The shape of the strain filter defines the quality of the elastogram: a narrow, low filter will cause elastographic artifacts such as image noise and low dynamic range to plague the elastogram. Thus, the design of the optimal strain filter for a given situation is of utmost importance to the production of quality elastograms.

The signal decorrelation parameter and shape of the strain filter vary with changes in the cross-correlation window length (resolution) used to obtain the time-delay. A family of strain filter curves can be obtained at different resolution levels. The effect of the window length parameter on signal decorrelation has not been analyzed in this paper. The dependence of the correlation coefficient on the finite window length used in the cross-correlation analysis is discussed in [21]. The expression for the effective correlation coefficient is presented as a product of the peak value of the correlation coefficient (used in this paper) and a derating factor. The derating factor accounts for the decay in the correlation coefficient with increased window lengths. The strain filter described in this paper is therefore more optimistic in its prediction of the strain estimator performance. In addition, the strain filters obtained at varying depths in tissue also would be different. The 2-D and non-stationary properties of the strain filter will be discussed in a later publication. It has also been demonstrated that under certain conditions, i.e. optimal multicompression with temporal stretching [23], the bandpass characteristic of the strain filter changes into a more desirable, high-emphasis characteristic.

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Appendix A

A. Combining $SNR_S$ and $SNR_\rho$ to obtain $SNR_C$

The expression for $SNR_C$ which is used to compute the post integration SNR and the thresholds in the ZZLB is presented here. The correlation coefficient is converted into an SNR measure [7], [18], that is given by:

$$SNR_\rho = \frac{\rho}{1 - \rho}.$$  \hspace{1cm} (A.1)

The variation of $SNR_\rho$ with a linear variation in $\rho$ is illustrated in Fig. 7. Note from (A-1) that for $\rho = 1$, $SNR_\rho = \infty$, dropping exponentially to an $SNR_\rho$ value of $22.46$ dB for $\rho = 0.93$. The composite value of the SNR, combining the contributions of $SNR_S$ and $SNR_\rho$, is given by [13], [14]:

$$SNR_C = \frac{SNR_S \cdot SNR_\rho}{1 + SNR_S + SNR_\rho}.$$  \hspace{1cm} (A.2)

This expression for $SNR_C$ incorporates both the electronic noise level and the decrease in $SNR$ caused by increase in signal decorrelation with strain. From (A.2) we observe that $SNR_C$ will always be bounded by the smallest value of either $SNR_S$ or $SNR_\rho$.

Appendix B

B. Lower Bound on the Variance of the Time Delay Estimator

The modified ZZLB derived by Weinstein and Weiss [13], [14] gives the tightest lower bound on the TDE variance. We present here the results for bandpass signals with
characterized by the constant variance level of \( (cT)^2/12 \), which corresponds to the variance of a random variable uniformly distributed between \(-sT/2 < \tau < sT/2\). In the threshold region, the TDE variance increases exponentially with the post integration SNR.

The thresholds given in (B.2) are computed for the following typical signal parameters: \( T = 1 \text{ mm } (1.3 \mu s), f_o = 5 \text{ MHz}, B = 3 \text{ MHz } (60\% \text{ bandwidth}), \text{ and } SNR_S = 40 \text{ dB} \). Assuming \( SNR_T \gg 40 \text{ dB} \), \( SNR_C \approx 40 \text{ dB} \). The numerical values of the thresholds \( \eta, \mu, \delta \), and \( \gamma \) and in (B-2) are 13.52, 0.78, 0.59, and 0.46 (22.6, \(-1.1, -2.3,\) and \(-3.4 \) when expressed in dB), respectively, at a post integration SNR value of 43.86 dB. The reader is referred to the papers by Weinstein and Weiss [13],[14] for a detailed description and derivation of the ZZLB and the thresholds for narrow and wide-band systems.

**References**


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